

(FILE 'HOME' ENTERED AT 08:54:47 ON 01 DEC 2003)

FILE 'MEDLINE, CAPLUS, BIOSIS, AGRICOLA' ENTERED AT 08:54:50 ON 01 DEC 2003

L1 55 S RPN11  
L2 33 S L1 AND UBIQUITIN  
L3 18 DUP REM L2 (15 DUPLICATES REMOVED)  
L4 4 S L3 AND SIC1

FILE 'STNGUIDE' ENTERED AT 08:56:16 ON 01 DEC 2003

FILE 'MEDLINE, CAPLUS, BIOSIS, AGRICOLA' ENTERED AT 08:59:52 ON 01 DEC 2003

L5 29 DUP REM L1 (26 DUPLICATES REMOVED)  
L6 4 S L5 AND (INHIBIT? OR MODIFY?)  
L7 133 S SIC1 AND UBIQUITIN  
L8 68 DUP REM L7 (65 DUPLICATES REMOVED)  
L9 659 S CULLIN AND UBIQUITIN  
L10 0 S L9 AND CU11  
L11 171 S L9 AND CUL1  
L12 1 S L1 AND JAB

=> s l1 and jam

L13 1 L1 AND JAM

L3 ANSWER 17 OF 18 MEDLINE on STN DUPLICATE 8  
 AN 2000414768 MEDLINE  
 DN 20372738 PubMed ID: 10913188  
 TI Evidence for separable functions of Srplp, the yeast homolog of importin alpha (Karyopherin alpha): role for Srplp and Stslp in protein degradation.  
 AU Tabb M M; Tongaonkar P; Vu L; Nomura M  
 CS Departments of Microbiology and Molecular Genetics and Biological Chemistry, University of California, Irvine, Irvine, California 92697-1700, USA.  
 NC GM35949 (NIGMS)  
 SO MOLECULAR AND CELLULAR BIOLOGY, (2000 Aug) 20 (16) 6062-73.  
 Journal code: 8109087. ISSN: 0270-7306.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 200008  
 ED Entered STN: 20000907  
 Last Updated on STN: 20030214  
 Entered Medline: 20000828  
 AB Srplp (importin alpha) functions as the nuclear localization signal (NLS) receptor in *Saccharomyces cerevisiae*. The srpl-31 mutant is defective in this nuclear localization function, whereas an srpl-49 mutant exhibits defects that are unrelated to this localization function, as was confirmed by intragenic complementation between the two mutants. **RPN11** and STS1 (DBF8) were identified as high-dosage suppressors of the srpl-49 mutation but not of the srpl-31 mutation. We found that Stslp interacts directly with Srplp in vitro and also in vivo, as judged by coimmunoprecipitation and two-hybrid analyses. Mutants of Stslp that cannot interact with Srplp are incapable of suppressing srpl-49 defects, strongly suggesting that Stslp functions in a complex with Srplp. STS1 also interacted with the second suppressor, **RPN11**, a subunit of the 26S proteasome, in the two-hybrid system. Further, degradation of Ub-Pro-beta-galactosidase, a test substrate for the **ubiquitin**-proteasome system, was defective in srpl-49 but not in srpl-31. This defect in protein degradation was alleviated by overexpression of either **RPN11** or STS1 in srpl-49. These results suggest a role for Srplp in regulation of protein degradation separate from its well-established role as the NLS receptor.

L3 ANSWER 15 OF 18 MEDLINE on STN DUPLICATE 6  
 AN 2002491539 MEDLINE  
 DN 22239942 PubMed ID: 12353037  
 TI A cryptic protease couples deubiquitination and degradation by the proteasome.  
 CM Comment in: Nature. 2002 Sep 26;419(6905):351-3  
 AU Yao Tingting; Cohen Robert E  
 CS Department of Biochemistry, University of Iowa, 51 Newton Road, Iowa City, Iowa 52242, USA.  
 SO NATURE, (2002 Sep 26) 419 (6905) 403-7.  
 Journal code: 0410462. ISSN: 0028-0836.  
 CY England: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 200210  
 ED Entered STN: 20020928  
 Last Updated on STN: 20021031  
 Entered Medline: 20021018  
 AB The 26S proteasome is responsible for most intracellular proteolysis in eukaryotes. Efficient substrate recognition relies on conjugation of substrates with multiple **ubiquitin** molecules and recognition of the polyubiquitin moiety by the 19S regulatory complex--a multisubunit assembly that is bound to either end of the cylindrical 20S proteasome core. Only unfolded proteins can pass through narrow axial channels into the central proteolytic chamber of the 20S core, so the attached polyubiquitin chain must be released to allow full translocation of the substrate polypeptide. Whereas unfolding is rate-limiting for the degradation of some substrates and appears to involve chaperone-like activities associated with the proteasome, the importance and mechanism of degradation-associated deubiquitination has remained unclear. Here we report that the POH1 (also known as **Rpn11** in yeast) subunit of the 19S complex is responsible for substrate deubiquitination during proteasomal degradation. The inability to remove **ubiquitin** can be rate-limiting for degradation in vitro and is lethal to yeast. Unlike all other known deubiquitinating enzymes (DUBs) that are cysteine proteases, POH1 appears to be a Zn(2+)-dependent protease.



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☐ 1: Mol Cell. 2001 Aug;8(2):439-48.

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**Selective degradation of ubiquitinated Sic1 by purified 26S proteasome yields active S phase cyclin-Cdk.****Verma R, McDonald H, Yates JR 3rd, Deshaies RJ.**

Howard Hughes Medical Institute, Division of Biology, California Institute of Technology, Pasadena, CA 91125, USA.

Selective degradation of single subunits of multimeric complexes by the ubiquitin pathway underlies multiple regulatory switches, including those involving cyclins and Cdk inhibitors. The machinery that segregates ubiquitinated proteins from unmodified partners prior to degradation remains undefined. We report that ubiquitinated Sic1 (Ub-Sic1) embedded within inactive S phase cyclin-Cdk (S-Cdk) complexes was rapidly degraded by purified 26S proteasomes, yielding active S-Cdk. Mutant proteasomes that failed to degrade Ub-Sic1 activated S-Cdk only partially in an ATP-dependent manner. Whereas Ub-Sic1 was degraded within approximately 2 min, spontaneous dissociation of Ub-Sic1 from S-Cdk was approximately 200-fold slower. We propose that the 26S proteasome has the intrinsic capability to extract, unfold, and degrade ubiquitinated proteins while releasing bound partners untouched. Activation of S-Cdk reported herein represents a complete reconstitution of the regulatory switch underlying the G1/S transition in budding yeast.

PMID: 11545745 [PubMed - indexed for MEDLINE]

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